

Article

Determination of the cervical vertebra maturation degree from lateral radiography

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Abstract: Many environmental and genetic conditions may modify jaws growth. In orthodontics, the right treatment timing is crucial. This timing is a function of the Cervical Vertebra Maturation (CVM) degree. Thus, determining the CVM is important. In orthodontics, the lateral X-ray radiography is used to determine it. Many classical methods need knowledge and time to look and identify some features to do it. Nowadays, Machine Learning (ML) and Artificial Intelligent (AI) tools are used for many medical and biological image processing, clustering and classification. This paper reports on the development of a Deep Learning (DL) method to determine directly from the images the degree of maturation of CVM classified in six degrees. Using 300 such images for training and 200 for evaluating and 100 for testing, we could obtain a 90% accuracy. The proposed model and method are validated by cross validation. The implemented software is ready for use by orthodontists.

Keywords: Classification, orthodontics, Cervical Vertebra Maturation, Machin Learning, Artificial Intelligence, Deep Learning

1. Introduction

1.1. Importance of the work and its interest for orthodontics community

Specialists in orthodontics are responsible for the treatment of dentofacial dysmorphisms, from different functional, genetical and morphological aetiologies. As a child or teenager is still growing, orthodontic treatment consists in a combination of orthodontics (about tooth position) and dentofacial orthopedics (about the guidance and stimulation of facial, maxilla and mandible growth in the three dimensions).

Many environmental and genetic conditions may induce upper or lower jaws lacks of growth. Classically, to handle a treatment properly, every etiological condition that can be modified or corrected, must be identified (diagnosis), normalized (treatment), and stabilized (retention). Specialists have to carefully examine and precisely analyze, all the medical, functional, clinical and radiographic data, in order to identify normal versus pathological conditions about tooth position, form or size, about lip, chin, cheeks, tongue and breathing functions, and about facial and jaws position and growing patterns. Adolescent orthodontic treatment also depends on proper management of jaws and facial growth, to allow a balanced jaws position, maximize the airway and improve the facial appearance. [1] Treatment planning in orthodontics depends on a systematic diagnosis and prognosis

Contemporary theories about craniofacial growth admit that the phenotype of the craniofacial complex is a result of a combination of genetic, epigenetic and environmental factors. The skeletal tissue of maxillomandibular complex is growing due to sutures and osteogenic cartilages proliferation depending on genetic, intrinsic and extrinsic environment. So facial growth can also be modified in amount and direction by extrinsic factors, including orthopedic and functional treatment. Thus,

34 quantify facial and, in particular, mandibular growth remaining, influences diagnosis, prognosis,
35 treatment goals and planning. Indeed, apart choosing the good appliance needed to change the rate
36 and direction of jaw growth, the right treatment timing is crucial. If high growth rate is about to
37 occur, orthopedic treatment may permit to correct jaws unbalanced, otherwise surgical correction of
38 the jaws shift will be considered. The success of a dentofacial orthopedic treatment is linked to the
39 determination of the best interventional frame (periods of accelerated or intense growth) to maximize
40 the chances to reach skeletal goals, with adapted methods and devices, in an optimized duration.

41 The most common dentofacial dysmorphism, is the skeletal class II, corresponding to a short
42 mandible. Study of normal mandibular growth and remodeling, has shown different ways of bone
43 formation, that can be stimulated by functional and orthopedics treatments, in particular condylar
44 growth responsible of 80% of the mandible growth. Numerous radiographic investigations have
45 established that condylar/mandibular growth follows similar growth curve than statural growth.[2]
46 This growth pattern is characterized by variations of growth rate in 4 stages: first a decrease of growth
47 velocity from birth to 6 years old, then minor midgrowth spurt around 6 to 8 years, followed by a
48 prepubertal plateau with decelerated growth rate, and finally the facial growth curve describe a peak
49 of growth velocity corresponding at the pubertal growth spurt, which coincides, precedes or follows
50 from 6 to 12 months the statural growth peak (controversial).[3] This spurt occurs approximately two
51 years earlier in girls than in boys.[4]

52 To estimate mandibular growth potential left, the patient must be localized on is growth curve,
53 and many biologic indicators have been proposed: increase in body height, menarche, breast and voice
54 changes, dental development and eruption, middle phalanx maturation of the third finger, maturation
55 of the hand and wrist, and cervical vertebral maturation. [3,7,8]

56 1.2. *The classical radiographic manual methods*

57 1.2.1. Hand-wrist radiograph method HWM:

58 The comparison method describes in the Atlas of Greulich et Pyle in 1959 or the Fishman's method
59 in 1982, permit to identify specific ossification stages occurring before, during, or after mandibular
60 growth peak, on left hand and wrist radiographs.[9,10] The hand wrist radiographs have been used
61 as a gold standard in the assessment of skeletal maturation for many decades, but presented several
62 issues as: the additional x-ray exposure, the time spending and experience required (even if a digital
63 software is now available [11]), and a sexual dimorphism and ethnic polymorphism in morphological
64 modifications.[12,13]

65 1.2.2. Vertebrae maturation CVM:

66 First who proposed to predict skeletal age and growth potential by cervical vertebrae maturation
67 (CVM) method is LAMPARSKI in 1972. Cervical vertebrae are available on the lateral cephalometric
68 radiographs, prescribed routinely by orthodontists for each patient diagnosis and treatment
69 planning.[14] He has used measurements of mandibular length on several annual lateral cephalograms
70 to describe individual mandibular growth curve, and correlated it with morphological description
71 of vertebrae morphology at each stage. This method were modified several times first by Hassel and
72 Farman (1995)[15], then twice by Baccetti et al. (2002 and 2005) for a more accurate assessment of
73 cervical maturation, by 6 stages identified by morphological changes in the C2,C3,C4 vertebral bodies
74 on a single lateral cephalogram, independently of patient gender. [16]

75 This last version is the most used nowadays to detect the mandibular growth spurt, as it shows
76 the best results in clinical applicability.[17]

77 As every single bones of the human body, vertebrae growth and present maturational changes
78 from birth to full maturity. Cervical vertebrae are the first seven pieces of the spinal column. Vertebral
79 growth in the cartilaginous layer of the superior and inferior surfaces of each vertebrae, involves
80 changes in size of vertebral bodies and shape of upper and lower borders of C2,C3,C4 vertebrae.

81 These changes have been described into 6 stages, correlating with morphological modifications of
 82 the vertebral shapes and estimated time lapse from the mandibular growth peak. Both visual and
 83 cephalometric appraisals of morphological changes have been proposed.

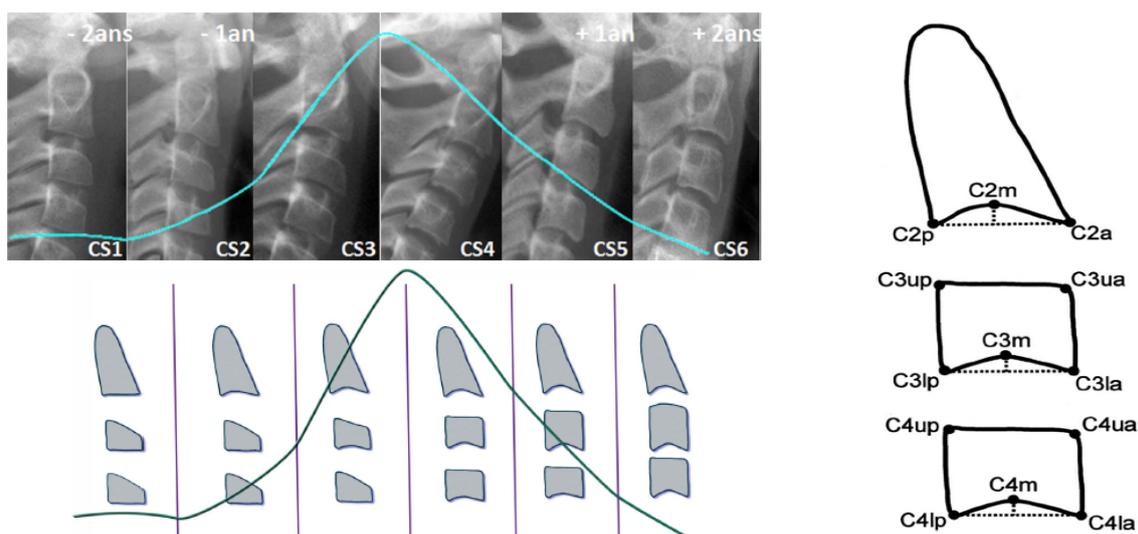


Figure 1. Left) CVM radiological and morphological stages superposed with Björk growth curve.[16],
 Right) Cephalometric landmarks for CVM stages determination.[1]

84 Visual analysis [1]:

- 85 • Cervical stage 1 (CS1) = 2 years before mandibular growth peak:
 86 Lower borders of C2 to C4 vertebrae are flat. C3 and C4 superior borders are tapered from
 87 posterior to anterior.
- 88 • Cervical stage 2 (CS2) = 1 year before mandibular growth peak:
 89 Lower border of C2 presents a concavity. Bodies of C3 and C4 are the same.
- 90 • Cervical stage 3 (CS3) = during the year of the mandibular growth peak:
 91 Lower borders of C2 and C3 present concavities. Vertebrae are growing so C3 and C4 may be
 92 either trapezoid or rectangular shape, as superior borders are less and less tapered.
- 93 • Cervical stage 4 (CS4) = 1 or 2 years after mandibular growth peak:
 94 Lower borders of C2, C3 and C4 present concavities. Both C3 and C4 bodies are rectangular with
 95 horizontal superior borders longer than higher.
- 96 • Cervical stage 5 (CS5) = 1 year after the end of mandibular growth peak:
 97 Still concavities of lower borders of C2, C3 and C4. At least one of C3 or C4 bodies are squared
 98 and spaces between bodies are reduced.
- 99 • Cervical stage 6 (CS6) = 2 years after the end of mandibular growth peak:
 100 The concavities of lower borders of C2 to C4 have deepened. C3 and C4 bodies are both square
 101 or rectangular vertical in shape (bodies higher than wide)

102 1.2.3. Cephalometric appraisals:

103 Using the landmarks illustrated on Figure 1(right), cephalometric analysis consists in the
 104 measurement of:

- 105 • The concavity depth of the lower vertebral border (estimated by the distance of the middle point
 106 (Cm) from the line connecting posterior to anterior points (Cip-Cla))
- 107 • The tapering of upper border of vertebral C3 and C4 bodies (estimated by the ratio between
 108 posterior and anterior bodies heights (Cup-CIp)/(Cua-Cla))
- 109 • The lengthening of vertebral bodies (estimated by the ratio between the bases length and anterior
 110 bodies borders height (CIp-Cla)/Cua-Cla)

111 Many researchers found this method as valid and reliable as hand and wrist Xray.[14] The cervical
112 vertebrae maturation stages have been demonstrated as a clinically useful maturation indicators for
113 evaluation of pubertal growth height and mandibular velocities [18,19,20], by correlation between
114 chronological age and cervical vertebrae maturation, between hand-wrist and cervical-vertebrae
115 maturation.[16,21,22,23]

116 Some studies underlined the need for association with other clinical assessments [24] in clinical
117 practice, and a good reliability in differentiating pre and post mandibular growth spurt periods.[25]

118 *1.3. The difficulties of the labeling task*

119 Specific training is provided to assess CVM stages reliably, and repeatably at a satisfactory
120 level.[26,27] Gabriel et al. minimized the risk of bias (radiographs without tracings, standardized
121 training to private practice orthodontists...) and observed a moderate intra and inter-observer
122 agreement (30 to 62% of cases). These results confirm the expertise required to proper determination
123 of CVM stage, and may be explained by the use of a qualitative method of assessment, and the lack in
124 detecting exceptional cases (individual variations in size and morphology, outside the norms defined
125 by the method). Moreover, for orthodontists, the cervical vertebrae area on the lateral cephalograms
126 is outside their expertise "visual field". They have poor general knowledge and experience about
127 vertebrae observation, as they focus on maxillomandibular bones and teeth at first glance. This would
128 have been a difficulty in the labeling task of our radiographs. All lateral radiographs have been
129 labeled by a radiologic technician, specialized in cephalometric tracing and over trained in CVM stages
130 agreement (3 years full time), using a standardized morphologic and cephalometric protocol. Intra
131 observer reproducibility must be estimated in further study.

132 *1.4. The need for automatization and the help which it brings*

133 Estimation of CVM stage represents only one single element influencing the patient orthodontic
134 treatment. The practitioner must master the entire clinical, functional, biomechanical and cephalometric
135 data analysis in order to define proper diagnosis and treatment goals and planning. Even in being
136 specialists, orthodontists require a very broad range of skills and a great deal of time for each patient
137 complete diagnosis. Considering that reproducibility of classifying CVM stages is superior at 98%
138 by trained examiners[1], automatization by expert eyes will provide time saving, efficiency, accuracy,
139 repeatability in treatment planning and patient care.

140 Few studies have presented software programs for cephalometric determination of C2,C3 and C4
141 vertebrae proportions according reference points marked manually on the image, and automatically
142 calculates the skeletal maturation stage. This computer-assisted analysis still depends on operator
143 experience.[28]. Padalino et al run a study comparing manual analysis of CVM stages and the analysis
144 performed by a dedicated software. It has shown a concordance of 94% between the two methods but
145 hand-tracing analysis was quicker of 28 seconds on average.[29]

146 Deep learning conventional neural networks have already been used to diagnose metabolic
147 disorders in pediatric endocrinology, in order to assess skeletal bone age on left hand-wrist radiographs.
148 Deep learning approach proposes better accuracy than conventional methods in processing the image
149 in less than 1 second. Our study aims to develop a fully automated deep learning assessment of CVM
150 stages on lateral cephalograms in orthodontics.

151 **2. Preprocessing of the data**

152 For this classification task, we had an image data base of 2000 X ray radiographic images. Each
153 image has a size of 2012x2012. These images are extracted from the patients files and are anonymized.

154 A selection of 600 images are studied and labeled by the experts in six classes (CVS1,...,CVS6).
155 These labeled data are divided in three sets of Training, Validation and Testing. We did different
156 division of the data: First, we had started by 300, 200 and 100, respectively for Training, Validation and

157 Testing. Then, we decided to divide them to 200, 200 and 200 and used Cross Validation technique by
 158 permutation of these sets.

159 Also, as these images are from the whole head, only a specific part of the image is usefull for
 160 this classification, we performed different preprocessing before feeding then to the DL input. In a
 161 preprocessing step, each original image is first cropped to the interesting part (Test1: size 488x488),
 162 then resized to (Test2: 244x244) or (Test3: 64x64) and after resizing to 244x244, they are Sobel filtered to
 163 enhance the contours of the image (Test 4). Figure 2 shows an example of these inputs.

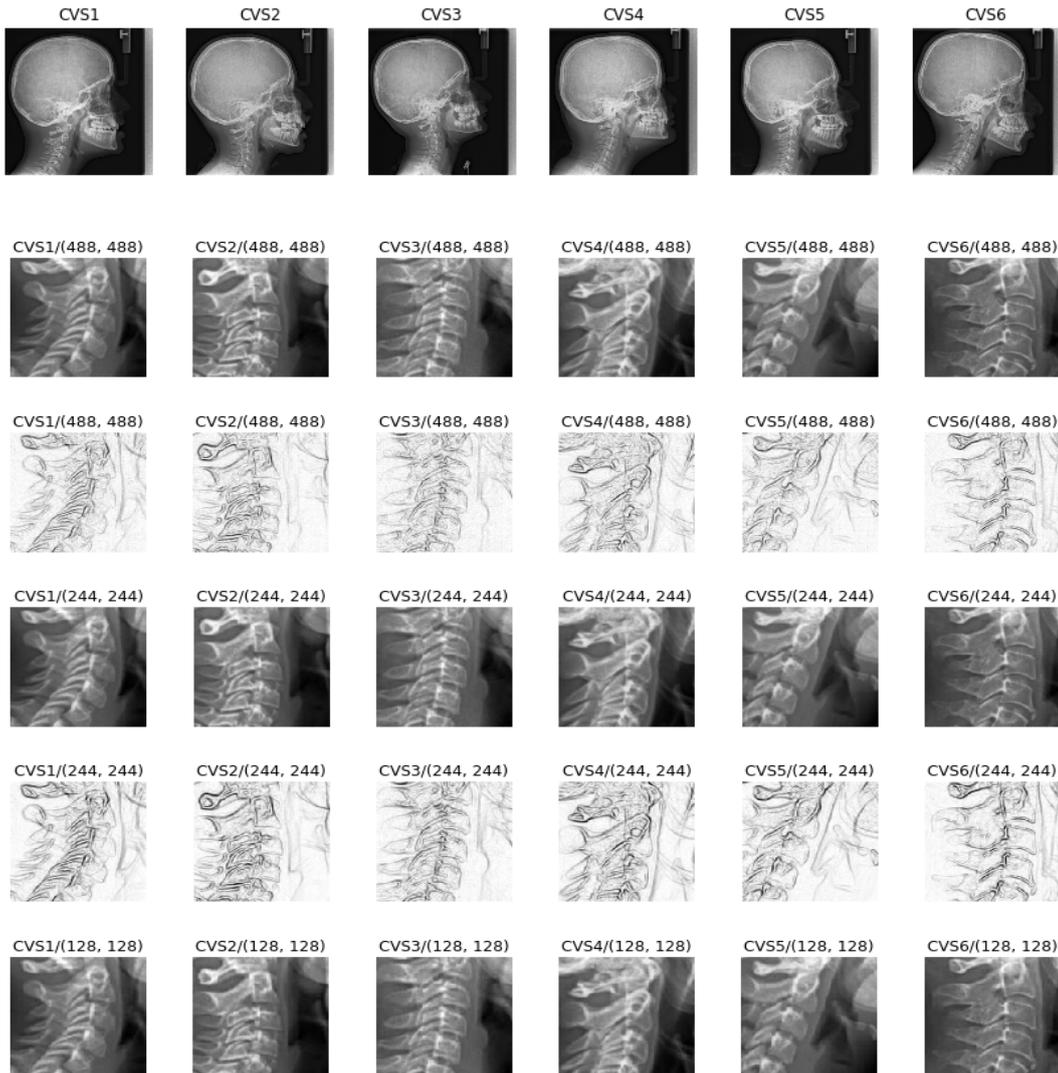


Figure 2. Originals and different preprocessing before training: a) Originals (2012x2020), b) test0: cropped images (488x488), c) test1: cropped and sobel edge detector filter (488x488), d) test2: cropped and resized (244x244), e) test3: cropped, resized and sobel edge detector filter (244x244), f) test4: cropped and resized (64x64)

164 3. Structure of the Deep Learning network

165 In a preliminary study, we used different Deep Learning network structures for this classification
 166 task and finally we selected a Deep Learning structure (like resnet) which is adapted for our task.

167 We considered different classical networks:

168 • Resnet :

169 Resnet was introduced in the paper "Deep Residual Learning for Image Recognition <<https://arxiv.org/abs/1512.03385>>". There are several variants with different output sizes, including
170 Resnet18, Resnet34, Resnet50, Resnet101, and Resnet152, all of which are available from
171 torchvision models. As our dataset is small, we used Resnet18 that we adapted in our case
172 for 6 classes.
173

174 • Alexnet:

175 Alexnet was introduced in the paper "ImageNet Classification with Deep Convolutional Neural
176 Networks
177 <[https://papers.nips.cc/paper/4824-imagenet-classification-with-deep-convolutional-neural-
178 networks.pdf](https://papers.nips.cc/paper/4824-imagenet-classification-with-deep-convolutional-neural-networks.pdf)> and was the first very successful CNN on the ImageNet dataset.

179 • VGG:

180 VGG was introduced in the paper "Very Deep Convolutional Networks for Large-Scale Image
181 Recognition <<https://arxiv.org/pdf/1409.1556.pdf>>". Torchvision offers eight versions of VGG
182 with various lengths and some that have batch normalizations layers.

183 • Squeezenet:

184 The Squeezenet architecture is described in the paper "SqueezeNet: AlexNet-level accuracy with
185 50x fewer parameters and <0.5MB model size", <<https://arxiv.org/abs/1602.07360>>. It uses a
186 different output structure than the other models mentioned here. Torchvision has two versions
187 of Squeezenet. We used version 1.0.

188 • Densenet:

189 Densenet was introduced in the paper "Densely Connected Convolutional Networks", <<https://arxiv.org/abs/1608.06993>>. Torchvision has four variants of Densenet. Here we used
190 Densenet-121 and modified the output layer, which is a linear layer with 1024 input features, for
191 our case.
192

193 • Inception v3:

194 Inception v3 was first described in "Rethinking the Inception Architecture for Computer Vision",
195 <<https://arxiv.org/pdf/1512.00567v1.pdf>>. This network is unique because it has two output
196 layers when training. The second output is known as an auxiliary output and is contained in the
197 AuxLogits part of the network. The primary output is a linear layer at the end of the network.
198 Note, when testing we only consider the primary output.

199 As it can be seen from on Figure 3, the structure of Deep Learning model is composed of an input
200 convolutional layer and three or four other convolutional nets (CNN) layers and a fully connected
201 of (32x32) to 6 classes. Each of the three CNNs is followed by a normalization, pooling and dropout
202 layers with different dropout coefficients.

203 The models are trained with different partitions of the images in Training, Validation and Testing
204 sets. The following figure shows 300 images which have been prepared for the training, then validated
205 on 200 images and saved to be used for testing step. A set of 120 images are used for testing step and
206 the average score was 80 percent.

207 4. Tools and Implementation

208 In this work, we used the following tools:

- 209 • SciKit-Learn: Data shuffling, Kmeans and Gaussian Mixture clustering, Principal Component
210 Analysis and performance metrics.
- 211 • Keras wth tensorflow backend: VGG16, VGG19 and ResNet50 convolution network models with
212 ImageNet weights

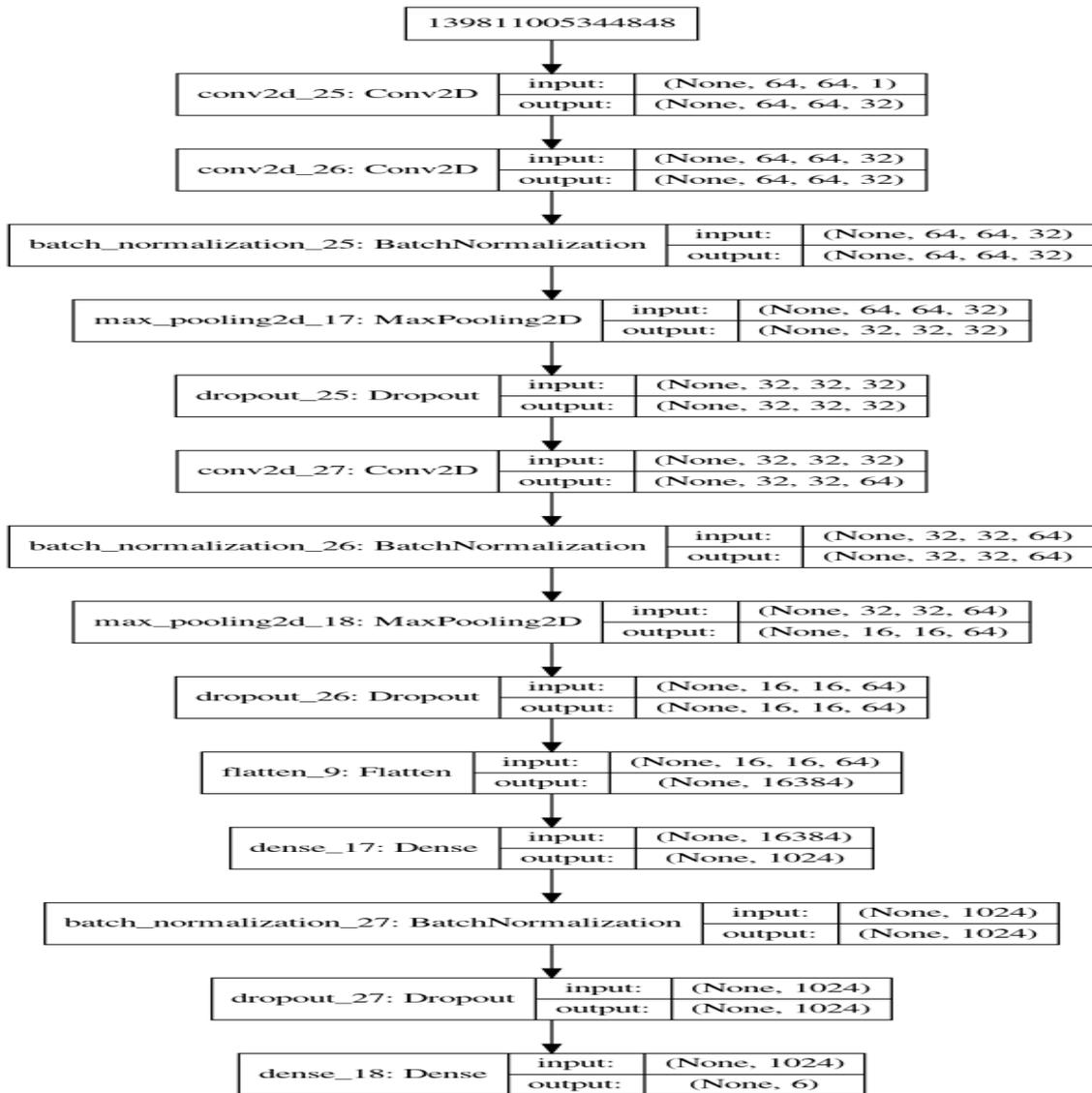


Figure 3. The structure of the proposed Deep Learning network

213 5. Prediction results

214 With implemented DL structure, we used 300 images for the training step, 200 images for the
 215 validation step and finally 150 images for the testing step. We had to fix a great number of parameters
 216 such as dropout rates, optimization algorithms, regularization parameters, etc.

217 The following figure shows the evolution of the Loss function and the accuracy as a function of
 218 the epoch numbers for one of these different tests.

219 Here, we show the prediction results obtained with different preprocessing of the data, both
 220 during the training and the testing

221 CVS1/ 0.83, CVS2/ 0.85, CVS3/ 0.72, CVS4/ 0.72, CVS5/ 0.79, CVS6/ 0.82

222 6. Conclusions

223 In this work, we developed and presented a specifically designed classification method for
 224 classifying the lateral radiographs of a great number of patients with the objective of determining the
 225 cervical vertebra maturation degree of bones, which is an important parameter for the orthodontists.

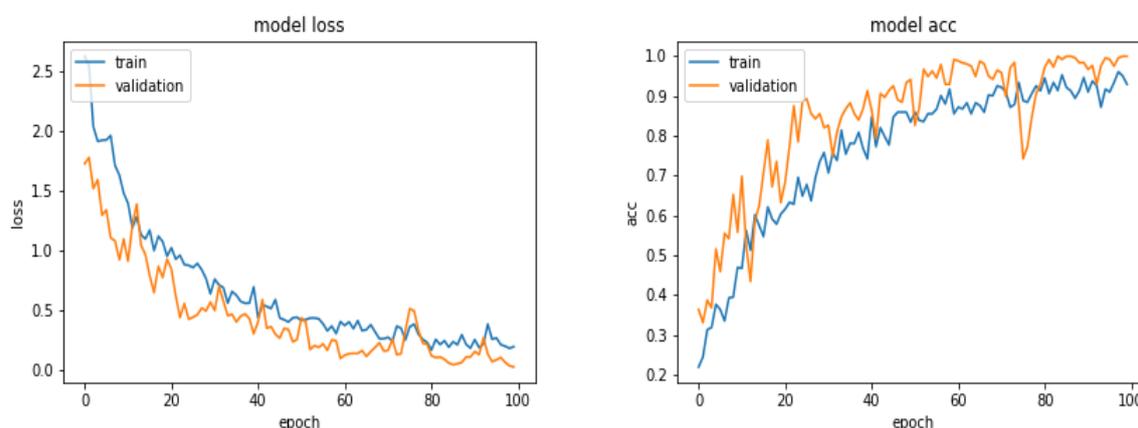


Figure 4. Evolution of the Loss function and the accuracy as a function of the epoch numbers.

226 The proposed Deep Learning classification method is particularly adapted for this task. In a first step,
 227 we used 300 labeled images for training, 200 for validation and hyper parameter tuning and finally 100
 228 for testing. Even if during the training and validation, we could obtain accuracies more than 95%, the
 229 accuracy for the testing images did not exceeded 85%. We think that with a greater number of training
 230 and validation images, this can be improved. Our plan is to use about 1000 images for training and
 231 1000 for testing in near future.

232 References

- 233 1. Baccetti T, Franchi L, McNamara JA. The Cervical Vertebral Maturation (CVM) Method for the Assessment
 234 of Optimal Treatment Timing in Dentofacial Orthopedics. *Semin Orthod.* sept 2005;11(3):119-29.
- 235 2. Patcas R, Herzog G, Peltomäki T, Markic G. New perspectives on the relationship between mandibular and
 236 statural growth. *Eur J Orthod.* févr 2016;38(1):13-21.
- 237 3. Moore RN, Moyer BA, DuBois LM. Skeletal maturation and craniofacial growth. *98(1):8.*
- 238 4. Hunter CJ. The correlation of facial growth with body height and skeletal maturation at adolescence.pdf.
 239 *Angle Orthod.* janv 1966;36(1):44-54.
- 240 5. Raberin M, Cozor I, Gobert-Jacquart S. Les vertèbres cervicales : indicateurs du dynamisme de la croissance
 241 mandibulaire? *Orthod Fr.* mars 2012;83(1):45-58.
- 242 6. Franchi L, Baccetti T, McNamara JA. Mandibular growth as related to cervical vertebral maturation and
 243 body height. *Am J Orthod Dentofacial Orthop.* sept 2000;118(3):335-40.
- 244 7. De Stefani A, Bruno G, Siviero L, Crivellin G, Mazzoleni S, Gracco A. évaluation radiologique de l'âge osseux
 245 avec la maturation de la phalange médiane du doigt majeur chez un patient orthodontique péripubertaire.
 246 *Int Orthod.* sept 2018;16(3):499-513.
- 247 8. Krisztina MI, A O, Réka G, Zsuzsa B. Evaluation of the Skeletal Maturation Using Lower First Premolar
 248 Mineralisation. *Acta Medica Marisiensis.* 1 déc 2013;59(6):289-92.
- 249 9. Pyle SI, Waterhouse AM, Greulich WW. Attributes of the radiographic standard of reference for the National
 250 Health Examination Survey. *Am J Phys Anthropol.* nov 1971;35(3):331-7.
- 251 10. Loder RT. Applicability of the Greulich and Pyle Skeletal Age Standards to Black and White Children of
 252 Today. *Arch Pediatr Adolesc Med.* 1 déc 1993;147(12):1329.
- 253 11. Bunch PM, Altes TA, McIlhenny J, Patrie J, Gaskin CM. Skeletal development of the hand and wrist: digital
 254 bone age companion—a suitable alternative to the Greulich and Pyle atlas for bone age assessment? *Skeletal
 255 Radiol.* juin 2017;46(6):785-93.
- 256 12. Srinivasan B, Padmanabhan S, Chitharanjan AB. Constancy of cervical vertebral maturation indicator in
 257 adults: A cross-sectional study. *Int Orthod.* sept 2018;16(3):486-98.
- 258 13. Shim J J, Bogowicz P, Heo G, Lagravère MO. Interrelationship and limitations of conventional radiographic
 259 assessments of skeletal maturation. *Int Orthod.* juin 2012;10(2):135-47.

- 260 14. O'Reilly MT, Yanniello GJ. 1988 O'REILLY Mandibular growth changes and maturation of cervical
261 vertebrae.pdf. *Angle Orthod.* 1988;
- 262 15. Hassel B, Farman AG. Skeletal maturation evaluation using cervical vertebrae. *Am J Orthod Dentofacial*
263 *Orthop.* janv 1995;107(1):58-66.
- 264 16. Elhaddaoui R, Benyahia H, Azaroual F, Zaoui F. Intérêt de la méthode de maturation des vertèbres cervicales
265 (CVM) en orthopédie dento-faciale: mise au point. *Rev Stomatol Chir Maxillo-Faciale Chir Orale.* nov
266 2014;115(5):293-300.
- 267 17. Jaqueira LMF, Armond MC, Pereira LJ, Alcântara CEP de, Marques LS. Determining skeletal maturation
268 stage using cervical vertebrae: evaluation of three diagnostic methods. *Braz Oral Res.* déc 2010;24(4):433-7.
- 269 18. Uysal T, Ramoglu SI, Basciftci FA, Sari Z. Chronologic age and skeletal maturation of the cervical vertebrae
270 and hand-wrist: Is there a relationship? *Am J Orthod Dentofacial Orthop.* nov 2006;130(5):622-8.
- 271 19. Hosni S, Burnside G, Watkinson S, Harrison JE. Comparison of statural height growth velocity at different
272 cervical vertebral maturation stages. *Am J Orthod Dentofacial Orthop.* oct 2018;154(4):545-53.
- 273 20. Perinetti G, Contardo L, Castaldo A, McNamara JA, Franchi L. Diagnostic reliability of the cervical vertebral
274 maturation method and standing height in the identification of the mandibular growth spurt. *Angle Orthod.*
275 juill 2016;86(4):599-609.
- 276 21. Mahajan S. Evaluation of skeletal maturation by comparing the hand wrist radiograph and cervical vertebrae
277 as seen in lateral cephalogram. *Indian J Dent Res.* 2011;22(2):309.
- 278 22. Sachan K, Tandon P, Sharma V. A correlative study of dental age and skeletal maturation. *Indian J Dent Res.*
279 2011;22(6):882.
- 280 23. Danaei SM, Karamifar A, Sardarian A, Shahidi S, Karamifar H, Alipour A, et al. Measuring agreement
281 between cervical vertebrae and hand-wrist maturation in determining skeletal age: Reassessing the theory
282 in patients with short stature. *Am J Orthod Dentofacial Orthop.* sept 2014;146(3):294-8.
- 283 24. Ball G, Woodside D, Tompson B, Hunter WS, Posluns J. Relationship between cervical vertebral maturation
284 and mandibular growth. *Am J Orthod Dentofacial Orthop.* mai 2011;139(5):e455-61.
- 285 25. Ballrick JW, Fields HW, Beck FM, Sun Z, Germak J. The cervical vertebrae staging method's reliability in
286 detecting pre and post mandibular growth. *Orthod Waves.* sept 2013;72(3):105-11.
- 287 26. Perinetti G, Caprioglio A, Contardo L. Visual assessment of the cervical vertebral maturation stages: A study
288 of diagnostic accuracy and repeatability. *Angle Orthod.* nov 2014;84(6):951-6.
- 289 27. McNamara JA, Franchi L. The cervical vertebral maturation method: « A user's guide ». *Angle Orthod.*
290 mars 2018;88(2):133-43.
- 291 28. Santiago RC, Cunha AR, Júnior GC, Fernandes N, Campos MJS, Costa LFM, et al. New software for cervical
292 vertebral geometry assessment and its relationship to skeletal maturation—a pilot study. *Dentomaxillofacial*
293 *Radiol.* févr 2014;43(2):20130238.
- 294 29. Padalino S, Sfondrini MF, Chenuil L, Scudeller L, Gandini P. Fiabilité de l'analyse de la maturité squelettique
295 selon la méthode CVM des vertèbres cervicales faite par un logiciel dédié. *Int Orthod.* dec 2014;12(4):483-93.